

Epithelial Barrier Dysfunction in Type 2 Inflammatory Diseases

This satellite symposium took place on 19th September 2019 as part of the 49th European Society for Dermatological Research (ESDR) Annual Meeting in Bordeaux, France

Chairperson:	Ignacio Dávila
Speakers:	Ignacio Dávila, ¹ Julien Seneschal, ² Sven-Erik Dahlén ³ <ol style="list-style-type: none">1. University Hospital, Salamanca, Spain2. Departments of Dermatology and Pediatric Dermatology, National Reference Center for Rare Skin Diseases, University of Bordeaux, France3. Unit for Experimental Asthma and Allergy Research at the National Institute for Environmental Medicine (NIEM) and the Centre for Allergy Research Karolinska Institutet (CfA), Stockholm, Sweden
Disclosure:	Prof Ignacio Dávila has acted as a speaker for ALK-Abello, Astra-Zeneca, Diater, GSK, Leti, Novartis, Sanofi, Stallergenes, and Teva; a consultant for Allergy Therapeutics, ALK-Abello, Astra-Zeneca, GSK, Merck, Novartis, Sanofi, and Stallergenes; and has received research support from Merck, and Thermofisher Diagnostics. Prof Julien Seneschal has acted as a speaker or consultant for Sanofi Genzyme, Lilly, AbbVie, LEO Pharma, Pierre Fabre, and has received institutional research support from Sanofi Genzyme, LEO Pharma, Lilly, Dermira, Pfizer, and AbbVie. Prof Sven-Erik Dahlén has acted as a speaker for, or collaborated in research with, Abbot Laboratories, Affibody AB, Altana Pharma, AstraZeneca, Aerocrine AB, Banyu Pharmaceutical, Bayer AG, Biolipox AB, Biovitrum AB, Boehringer Ingelheim, Cayman Chemical, Centocor Biotech, Genentech, GlaxoSmithKline, Johnson & Johnson, Kyorin Pharmaceutical, Meda AB, Merck & Co, Nigaard Pharma, Novartis, ONO Pharmaceutical, Pharmaxis, Regeneron Pharmaceuticals, RSPR Incentive AB, Sanofi SA, Schering-Plough Corporation, UCB, and Teva Pharmaceutical Industries Ltd.
Acknowledgements:	Writing assistance was provided by Nicola Humphry, Nottingham, UK.
Support:	The symposium and publication of this article were funded by Sanofi Genzyme.
Citation:	EMJ Dermatol. 2019;7[1]:44-51.

Meeting Summary

This satellite symposium took place during the 49th annual meeting of the European Society for Dermatological Research (ESDR). Prof Dávila began the symposium by describing the immunology behind Type 2 inflammation as a complex interaction between environmental factors, immune response, and barrier dysfunction. He explained that the principal cells participating in innate Type 2 immunity are Type 2 innate lymphoid cells (ILC2), eosinophils, basophils, and mast cells, and that Th2 lymphocytes, dendritic cells (DC), and their main cytokines (IL-4, IL-5, and IL-13) comprise the adaptive arm of the Type 2 immune response and are essential in IgE-mediated reactions. Prof Seneschal followed by explaining that Type 2 inflammation in atopic dermatitis (AD) is a combination of immune and epidermal barrier components influenced by genetic and environmental factors. Epidermal barrier proteins are expressed in lower levels in AD, and other proteins are also dysregulated, disrupting tight junctions. Both lesional and nonlesional skin in patients with AD show

epithelial barrier dysfunction, and inflammation can lead to a vicious cycle of itching and damage. Prof Dahlén concluded the meeting by explaining that airway inflammation is one of the major factors involved in Type 2 asthma, and this can be driven by an allergic route, involving mast cells, or a nonallergic route, involving ILC2. Inflammatory cytokines also increase mucus production, one of the main causes of asthma-related death. Recent studies of asthma immunology have suggested that ILC2 are subject to feedback modulation by prostaglandin D2 (PGD2), and that both IL-4 and IL-13 are involved in hyper-responsiveness in asthmatic lung tissue.

The Immunology of Type 2 Inflammation

Professor Ignacio Dávila

Type 2 immunity evolved from a dialogue between the immune system and microbes. This form of immunity confers protection against bacteria, viruses, and parasites such as helminths, but also promotes allergic inflammation, forming the basis of AD.

Type 2 immunity involves the activation and recruitment of immune cells, and the release of inflammatory cytokines.¹ Helminths trigger the ‘alarmins’ IL-25, IL-33, and thymic stromal lymphopoietin (TSLP), which activate Type 2 innate lymphoid cells (ILC2). ILC2 activate and amplify other Type 2 cells such as Th2 cells and eosinophils, and release Type 2 cytokines (for example, IL-4, IL-5, and IL-13).¹

Inflammation resulting from Type 2 immunity is a complex interaction between environmental factors, immune response (both innate and acquired), and barrier dysfunction.² Barrier dysfunction is a core feature of Type 2 inflammatory diseases; in asthma, respiratory viruses and allergens penetrate the epithelium causing an infiltration of immune cells and release of IL-4, IL-5, and IL-13, which disrupt bronchial epithelial tight junction proteins further.³⁻⁵ In AD, environmental allergens pass through the dermal barrier, encouraging the infiltration of immune cells to the area, and subsequent disruption of the stratum corneum and/or tight junctions.³

Proteases, endotoxins, and β -glucans can destroy tight junctions, leading to the production of NF- κ B and reactive oxygen species, and the release of IL-33, IL-25, and danger signals such as uric acid.⁶ These signals can act on immature DC (iDC) and ILC2 cells, causing inflammation.

Some nonproteolytic allergens and genetic polymorphisms in genes such as *SPINK5* can also induce barrier defects, facilitating the penetration of allergens and giving them increased access to DC, as well as the sensitisation of Th2 cells.⁶

Early immune responses are driven by ILC2. Following activation from the epithelium, they release IL-9, IL-13, and IL-5, causing goblet-cell hyperproliferation, subepithelial fibrosis, migration of eosinophils, and subsequent TGF- β production, and smooth muscle remodelling.⁶⁻⁹ ILC2 cells work through complex networks to stimulate the activation and proliferation of diverse immune cells, including macrophages, eosinophils, B cells, DC, and T cells, resulting in the production of prostaglandin D2 and other cytokines.¹⁰ Alarmins can also activate DC, which in turn are able to capture allergens and migrate to regional lymphatic nodes. There, they present antigens to naïve T cells, inducing a Th2 response.

Eosinophils are produced in the bone marrow, and display receptors for Ig, IL, and many other cytokines. These cells can produce molecules that affect nerves, such as prostaglandins, chemokines, and growth factors.^{11,12} Eosinophils also perform multiple immunomodulatory functions, including the activation of natural helper cells (NHC), macrophages, and Th2 cells, and the proliferation of T cells and neutrophils, resulting in the release of histamine and mast cell mediators.¹²

Mast cells and basophils both express receptors for IgE (high-affinity IgE receptor [Fc ϵ RI]), but priming factors differ, with mast cells primed by IL-4 and IL-6, and basophils primed by IL-3, IL-5, and IL-33. Each cell produces its own mediators, including IL-5 from mast cells, and IL-4 from basophils, as well as some common mediators, such as histamine and granzyme B.¹³ The mast cell network is very important for the Type 2 immune response. It connects mast cells with nerves, where they have a bidirectional influence through TNF and PGD2, causing irritation. The

release of mast-cell granules recruits leukocytes to the endothelium, and secretion of TNF stimulates proliferation of T cells causing lymph node hypertrophy.¹⁴ The role of basophils in the Type 2 immune response was long ignored, but peripheral basophils produce IL-4, facilitating Th2 differentiation following stimulation through the major histocompatibility complex class II. Basophils can also display the major histocompatibility complex class II molecules on their surface through trogocytosis, to stimulate Th2 differentiation.¹⁵ Mast cells and basophils perform different roles in allergic inflammation, with basophils particularly significant in IgE-mediated chronic allergic inflammation and IgE-independent asthma.¹⁶

Asthma is a condition caused by Type 2 inflammation in the airways.¹⁷ Epithelial cells release IL-25 which stimulates DC, activating Th2 cells through IL-4.¹⁷ Th2 cells activate B cells, releasing IgE, which later occupies mast cell receptors inducing the release of histamine and cytokines, causing smooth muscle contraction.¹⁷ Th2 cells also activate eosinophils through the release of IL-5, resulting in activation of the epithelium and consequent mucus production.¹⁷ Some types of Th1 cells are also involved in asthma, through the TNF- α /interferon gamma (IFN- γ)-mediated activation of neutrophils.¹⁷ The major Type 2 cytokines involved in asthma are IL-4, IL-5, and IL-13, all produced by Th2 and ILC2 cells.¹⁸ The release of these cytokines produces a multitude of effects on the airways, including smooth muscle contraction, hyperproduction of mucus, barrier disruption, tissue remodelling, and fibrosis.¹⁸

AD is a result of Type 2 inflammation in the skin, involving the activation of ILC2 cells through cytokines IL-25 and IL-33, followed by itching produced by IL-31 from Th2 cells.¹⁹ In the acute stage there is an increase in DC and Th2 activation, along with dilation of the vasculature, which leads to Th1 involvement and the release of IL-17, IL-22, and IFN- γ .¹⁹ The major cytokines involved in AD are IL-4 and IL-13, which together result in Th2 expansion, B cell class-switching and IgE production, increased vascular adhesion and permeability, production of chemoattractants, and stimulation of the itch-scratch cycle.²⁰⁻²⁶

In summary, Type 2 immunity has important functions in the body, such as defence against

parasites and venoms. It has three main components: the epithelial barrier, innate immunity, and acquired immunity. The principal cells participating in innate Type 2 immunity are ILC2 cells, eosinophils, basophils, and mast cells. Th2 lymphocytes, dendritic cells, and their main cytokines (IL-4, IL-5, and IL-13) comprise the adaptive arm of the Type 2 immune response and are essential in IgE-mediated reactions.

Epithelial Barrier Dysfunction and Type 2 Inflammation in Atopic Dermatitis

Professor Julien Seneschal

Healthy skin functions as a physical, permeability, and antimicrobial barrier against exogenous molecules or antigens, and maintains the internal environment.^{27,28} The terminally differentiated layer of the skin, the stratum corneum, consists of corneocytes embedded in a water-repellent, lipid-rich matrix that prevents transepidermal water loss and allergen absorption.^{27,29-31} Antimicrobial peptides, including β -defensin and cathelicidin, prevent colonisation and invasion by pathogenic microbes.³² Tight junctions form a structural barrier by sealing intracellular spaces and are often disrupted in AD.^{27,31,32} The plasma membrane of corneocytes is replaced by an insoluble protein structure known as the cornified cell envelope, and this forms a scaffold for lipid matrix attachment.^{29,33} Filaggrin, involucrin, and loricrin are important structural proteins in the cornified cell envelope, with filaggrin binding intracellular keratin fibres, and degrading into natural moisturising factor to maintain skin hydration and low pH.^{32,34-36}

The pathophysiology of AD has a multifactorial aetiology of immune and epidermal barrier components influenced by genetic and environmental factors.³⁷ Genetic factors can predispose individuals to AD, and include dysregulations of thymic stromal lymphopoietin (TSLP), IL-4/IL-13, toll-like receptor 2 (TLR2), IgE/Fc ϵ RI, filaggrin, serine protease inhibitor of the Kazal type (SPINK), and hornerin. Environmental factors can include allergy sensitisation, dryness, scratching, phototoxicity, and exposure to microbes or toxins which disrupt the skin barrier.

AD histology is characterised by marked spongiosis, which is vesicle formation from intraepidermal fluid, parakeratosis, the retention of nuclei in the stratum corneum, and subtle vacuolar and dermal-epidermal interface changes.^{38,39} Epidermal barrier proteins such as filaggrin are expressed in lower levels in AD. Although mutations in filaggrin have been observed, they are neither necessary nor sufficient for the development of AD, suggesting that other factors are involved.^{36,40} AD is associated with defects in the stratum corneum which result from filaggrin deficiency, including fewer keratohyalin granules, downregulated filaggrin degradation enzymes, dysregulated acidification pathways, and higher exposure to *Staphylococcus aureus*, herpes simplex virus Type 1 (HSV-1), and other allergens.⁴¹ Dysregulation of claudin-1 in AD causes disruption to tight junctions, which results in the elongation of Langerhans cell dendrites into the epidermal layer, encouraging antigen sensitisation.²⁷

In patients with AD, even normal-appearing skin has barrier defects, making these individuals more sensitive to environmental factors, *S. aureus*, and scratching.^{8,18,19} One of the characteristics of AD is the abnormal microbiome of the skin, with increased colonisation with *S. aureus*.⁴² *S. aureus* induces epithelial barrier disruption through toxin production and inflammation, resulting in increased protease activity. A recent study by Williams et al.⁴³ found that the introduction of *S. hominis* to the skin can inhibit *S. aureus* activity, preventing skin barrier disruption and inflammation.

The genetic expression profile in patients with AD, the 'AD transcriptome', shows that genes associated with Type 2 inflammation are upregulated in AD. However, since the transcriptome is similar in both lesional and nonlesional skin, there must also be a molecular aspect to the inflammation in lesional skin.⁴⁴ Inflammation in AD involves CD4+ and CD8+ T cells in both the dermis and epidermis, with the CD4+ cells secreting Type 2 cytokines IL-4 and IL-13, but again, this profile is similar between lesional and nonlesional skin.⁴⁵ AD is associated with increased expression of IFN γ , IL-13, and IL-22 in the skin compared to healthy individuals, and these cytokines are most abundant in lesional skin.⁴⁴ The expression levels of Type 2 cytokines IL-4, IL-5, IL-10, IL-13, and IL-31, in particular, correlate with disease activity in skin biopsies

patients with AD.⁴⁶ These Type 2 cytokines have been shown to downregulate epidermal barrier proteins filaggrin and hornerin,⁴⁷ and increase TSLP production, inducing spongiosis.⁴⁸ Type 2 inflammation induces defects in the epidermal barrier, increasing skin permeability and the cutaneous innate immune response. IL-4 and IL-13 result in epidermal thickening and disturb the expression of tight junction proteins such as occluding,⁴⁹ as well as exacerbating the itch-scratch cycle by sensitising neurons to pruritogens IL-31, TSLP, and histamines.²⁵

In summary, alteration of the epidermal barrier is an important feature of AD, promoting inflammation in both lesional and nonlesional skin associated with increased Type 2 proinflammatory cytokines IL-4 and IL-13. These cytokines induce defects in the epidermal barrier, leading to a vicious cycle that needs to be blocked.

Epithelial Barrier Dysfunction and Type 2 Inflammation in Asthma

Professor Sven-Erik Dahlén

It is an exciting time for asthma and respiratory disease, with many new therapeutic treatment strategies coming to market, and many more in the pipeline. In addition to new biologics, there are also new molecular targets and drug combinations, some with remarkable effects. Treatments for asthma have evolved over the last 100 years; from adrenaline, oral steroids, theophylline, and inhaled β 2-agonists, through inhaled anticholinergics and steroids, long-acting drugs, and sodium cromoglycate, to anti-IgE, antileukotrienes, anti-IL-5, and anti-IL-4R α .⁵⁰

Asthma Pathobiology

Asthma can be described as a condition whereby airways constrict too much, too often, and too easily, resulting in impaired lung physiology and quality of life. A bronchoscopy of patient with asthma, 3 hours after a bronchoprovocation challenge, will show that although lung function may have recovered, severe inflammation and oedema remain, and the airway is still quite narrow due to smooth muscle constriction. Pathologically, asthma has four components:

airway hyper-responsiveness, airway remodelling, airway inflammation (often eosinophilic with increased fractional exhaled nitric oxide [FENO]), and bronchoconstriction.

Airway hyper-responsiveness can be measured by airway reactivity (lung function) to inhaled methacholine, or histamine. In patients with severe asthma, lung function decreases significantly after a bronchoprovocation challenge with even low doses of methacholine/histamine, and these patients can be described as 'super responders'. Current treatments for asthma are poorly effective against this problem. Airway remodelling in asthma includes epithelial dysfunction; furthermore, asthmatic airways demonstrate increasing thickness of the epithelium and reticular basement membrane, and goblet cell hyperplasia as asthma severity increases.⁵¹ In asthmatic patients, external factors such as viruses, pollutants, and allergens are able to cross the dysfunctional epithelial barrier and trigger a Type 2 response. There are two major routes to Type 2 eosinophilic inflammation in the airway: the allergic route, which involves mast cells; and the nonallergic (innate) route, which involves ILC2 cells rarely seen in healthy lung tissue.² As well as eosinophilic inflammation, mucus production is stimulated by IL-13 and IL-9, and many asthma deaths are caused by mucus suffocation.⁵²

Bronchoconstriction in asthma can be triggered by antigens, which bind to IgE receptors on the surface of mast cells and induce secretion of histamine, cysteinyl leukotrienes, and prostanoids, causing contraction of smooth muscle in the airway.⁵³ In isolated human small bronchi, challenge with anti-IgE can mimic allergen exposure and result in bronchoconstriction. This reaction can be blocked with a combination of the thromboxane receptor antagonist SQ-29,548, the H1 antagonist mepyramine, and the leukotriene synthesis inhibitor MK-886.⁵³ This shows the dependence on the mast cell mediators for the immediate IgE-dependent reaction.

Biomarkers of Type 2 Asthma

The main biomarkers of Type 2 inflammation in asthma are eosinophils in the blood and sputum, FENO, IgE, and periostin. Dr Morrow Brown identified the important role of eosinophils in

chronic asthma in 1958, and this association has become more firmly established in the past 25 years.⁵⁴ Monitoring the eosinophilic content of sputum to guide treatment choices has been shown to reduce the severity of cumulative asthma exacerbations.⁵⁵

FENO levels relate to eosinophilic inflammation and can be used as another marker of Type 2 inflammation.^{56,57} When FENO is measured during repeated low-dose allergen challenges, it shows a clear sensitivity to steroid treatment.⁵⁸ A recent Phase III trial of dupilumab, a monoclonal antibody that inhibits IL-4 and IL-13, used FENO to predict patient response, demonstrating that patients with higher levels of FENO responded better to treatment.⁵⁹ U-BIOPRED (Unbiased BIOMarkers in PREDiction of respiratory disease outcomes) is a large, ongoing study which aims to identify novel biomarkers in severe asthma. A specialised 'breathomics' device is being used to recognise organic compounds in exhaled air of patients with severe asthma or mild/moderate asthma, and in healthy controls.⁶⁰ IgE still has a role as a biomarker of asthma, but periostin is being used less due to variable research findings.

Pivotal Role for Prostaglandin D2 in Type 2 Innate Lymphoid Cell Function

The nonallergic (innate) route to eosinophilic airway inflammation involves the stimulation of ILC2 cells.² ILC2 cells stimulated by alarmins such as IL-2, IL-25, IL-33, and TSLP secrete PGD2, and inhibition of PGD2 synthesis by either a nonsteroidal anti-inflammatory drug or an experimental drug results in reduced production of IL-5 and IL-13, suggesting that endogenously produced PGD2 is necessary for ILC2 cytokine production.⁶¹ This suggests ILC2 cells are subject to feedback modulation by PGD2 via the chemoattractant receptor-homologous molecule receptor.⁶³

In summary, the pathobiology of Type 2 asthma is driven by the inflammatory cytokines IL-4, IL-5, and IL-13 that are secreted in reactions involving Th2 cells, mast cells, eosinophils, and ILC2 cells. Although Type 2 asthma responds to inhaled steroids, this is not sufficient to provide optimal control in more severe cases. Eosinophils, FENO, and total IgE are currently used to indicate Type 2 inflammation in patients with asthma, and urinary LTE4 and PGD2 metabolites show promise as novel biomarkers.

Knowledge Gaps in Type 2 Inflammatory Diseases (Panel Discussion)

Professor Ignacio Dávila, Professor Julien Seneschal, and Professor Sven-Erik Dahlén

Q: Is Th2 inflammation equally important in both asthma and atopic dermatitis, and does it occur before or after the development of barrier defects?

Prof Dahlén indicated that 30 years ago, damaged epithelium was considered to be critical in the development of asthma, but that more recently it has been considered to be more of a functional consequence of increased levels of alarmins and PGE2 rather than damage per se. Prof Seneschal said that for AD, both biological defects and inflammation are important, and promote each other. The epidermal layer is important for the production of alarmins and many other cytokines. He noted that smooth muscle contraction is one of the main symptoms in asthma but not in AD, so perhaps the response of smooth muscle to specific cytokines may explain the differences between asthma and AD. Prof Seneschal replied that in asthma, 2–3 days' incubation of airways with IL-4 and IL-13 can transform the epithelium into an asthmatic phenotype. Many cytokines are important to modify immune cells and affect smooth muscle, but the main effectors of the contraction are histamine and prostaglandins.

Q: Would restoring the epithelial barrier in atopic dermatitis be useful in the prevention of atopic dermatitis, for example, through moisturisation?

Prof Dahlén said that healthcare providers try to educate patients to apply moisturising creams. There has been some encouraging data for the use of creams in the prevention of AD in newborn babies, but in general, skin moisturisation is important to prevent new skin defects and

flares in AD patients, in combination with anti-inflammatory medication.

Q: Type 2 inflammation is prominent in atopic dermatitis, but other types of inflammation are also important. In asthma, are Th2 cells more prominent than they are in atopic dermatitis?

Prof Dahlén replied that the Th2 paradigm is a huge simplification, and that severe asthma does seem to involve some neutrophilic inflammation, though it is unclear whether this is steroid-induced, or the result of bacteria. Efforts are being made through molecular phenotyping to identify new biomarkers; many different inflammatory subtypes for asthma can now be identified, but it is unclear how many are clinically relevant and can be independently targeted. Prof Seneschal said that in AD, patients could be grouped as IgE sensitive or insensitive, but the reasons for these differences can be very hard to understand.

CONCLUDING REMARKS

Professor Ignacia Dávila

In regard to the presented discussion, it would appear that although there is a Type 2 inflammatory basis to both AD and asthma, these conditions are not always associated, and do not always respond to the same treatments. The Type 2 Innovation Grant,⁶⁴ established in 2018, supports independent, novel, and innovative research projects designed to advance knowledge in the field of Type 2 immune-mediated diseases, with relevance to clinical practice. It targets not-for-profit organisations established in the European Economic Area and provides grants to research that is not related to investigational or marketed medicines. Applications are reviewed according to predefined criteria by independent international experts. Prof Seneschal is part of the expert dermatology panel established in 2019, and Prof Dahlén is on the respiratory panel. This innovation grant is proving a successful initiative, with 502 users, 252 active applications, 81 applications submitted to experts for review, and 8 selected projects thus far.

References

- Gazzinelli-Guimaraes PH, Nutman TB. Helminth parasites and immune regulation. *F1000Res*. 2018;7:1685.
- Lambrecht BN, Galli SJ. SnapShot: Integrated Type 2 immune responses. *Immunity*. 2015;43(2):408.
- Schleimer RP, Berdnikovs S. Etiology of epithelial barrier dysfunction in patients with Type 2 inflammatory diseases. *J Allergy Clin Immunol*. 2017;139(6):1752-61.
- Saatian B et al. Interleukin-4 and interleukin-13 cause barrier dysfunction in human airway epithelial cells. *Tissue Barriers*. 2013;1(2):e24333.
- Sugita K et al. Type 2 innate lymphoid cells disrupt bronchial epithelial barrier integrity by targeting tight junctions through IL-13 in asthmatic patients. *J Allergy Clin Immunol*. 2018;141(1):300-10.
- Hammad H, Lambrecht BN. Barrier epithelial cells and the control of Type 2 immunity. *Immunity*. 2015;43(1):29-40.
- Janeway CA et al., *Immunobiology (2001) 5th edition*, New York: Garland Science.
- Wynn TA. Type 2 cytokines: Mechanisms and therapeutic strategies. *Nat Rev Immunol*. 2015;15(5):271-82.
- Doherty TA, Broide DH. Airway innate lymphoid cells in the induction and regulation of allergy. *Allergol Int*. 2019;68(1):9-16.
- van Rijjt L et al. Type 2 innate lymphoid cells: At the cross-roads in allergic asthma. *Semin Immunopathol*. 2016;38(4):483-96.
- O'Sullivan JA, Bochner BS. Eosinophils and eosinophil-associated diseases: An update. *J Allergy Clin Immunol*. 2018;141(2):505-17.
- Sastre B et al. Eosinophils: Old players in a new game. *J Investig Allergol Clin Immunol*. 2018;28(5):289-304.
- Varricchi G et al. Human mast cells and basophils-How are they similar how are they different? *Immunol Rev*. 2018;282(1):8-34.
- Stassen M et al. Mast cells within cellular networks. *J Allergy Clin Immunol*. 2019;144(4S):46-54.
- Miyake K, Karasuyama H. Emerging roles of basophils in allergic inflammation. *Allergol Int*. 2017;66(3):382-91.
- Kubo M. Mast cells and basophils in allergic inflammation. *Curr Opin Immunol*. 2018;54:74-9.
- Pelaia G et al. Cellular mechanisms underlying eosinophilic and neutrophilic airway inflammation in asthma. *Mediators Inflamm*. 2015;2015:879783.
- Gandhi NA et al. Targeting key proximal drivers of Type 2 inflammation in disease. *Nat Rev Drug Discov*. 2016;15(1):35-50.
- Noda S et al. The translational revolution and use of biologics in patients with inflammatory skin diseases. *J Allergy Clin Immunol*. 2015;135(2):324-36.
- Chen L et al. VCAM-1 blockade delays disease onset, reduces disease severity and inflammatory cells in an atopic dermatitis model. *Immunol Cell Biol*. 2010;88(3):334-42.
- Palomares O et al. Mechanisms of immune regulation in allergic diseases: The role of regulatory T and B cells. *Immunol Rev*. 2017;278(1):219-36.
- Weidinger S et al. Atopic dermatitis. *Nat Rev Dis Primers*. 2018;4(1):1.
- Rerknimitr P et al. The etiopathogenesis of atopic dermatitis: Barrier disruption, immunological derangement, and pruritus. *Inflamm Regen*. 2017;37:14.
- Brunner PM et al. The immunology of atopic dermatitis and its reversibility with broad-spectrum and targeted therapies. *J Allergy Clin Immunol*. 2017;139(4S):65-76.
- Oetjen LK et al. Sensory neurons co-opt classical immune signaling pathways to mediate chronic itch. *Cell*. 2017;171(1):217-28.
- Patel KD. Eosinophil tethering to interleukin-4-activated endothelial cells requires both P- selectin and vascular cell adhesion molecule-1. *Blood*. 1998;92(10):3904-11.
- De Benedetto A et al. Tight junction defects in patients with atopic dermatitis. *J Allergy Clin Immunol*. 2011;127(3):773-86.
- Bolognia J et al, "Anatomy and Physiology," Vandergrill T, Bergstresser P (eds.), *Dermatology (2012) 3rd edition*, China: Elsevier Limited.
- Brogden NK et al. The emerging role of peptides and lipids as antimicrobial epidermal barriers and modulators of local inflammation. *Skin Pharmacol Physiol*. 2012;25(4):167-81.
- Harder J et al. The skin surface as antimicrobial barrier: Present concepts and future outlooks. *Exp Dermatol*. 2013;22(1):1-5.
- Clausen M-L et al. In vivo expression of antimicrobial peptides in atopic dermatitis. *Exp Dermatol*. 2016;25(1):3-9.
- Simpson CL et al. Deconstructing the skin: Cytoarchitectural determinants of epidermal morphogenesis. *Nat Rev Mol Cell Biol*. 2011;12(9):565-80.
- Candi E et al. The cornified envelope: A model of cell death in the skin. *Nat Rev Mol Cell Biol*. 2005;6(4):328-40.
- Kabashima K. New concept of the pathogenesis of atopic dermatitis: Interplay among the barrier, allergy, and pruritus as a trinity. *J Dermatol Sci*. 2013;70(1):3-11.
- Brown SJ, McLean WHI. One remarkable molecule: Filaggrin. *J Invest Dermatol*. 2012;132(3 Pt2):751-62.
- Irvine AD et al. Filaggrin mutations associated with skin and allergic diseases. *N Engl J Med*. 2011;365(14):1315-27.
- Eyerich K, Novak N. Immunology of atopic eczema: Overcoming the Th1/Th2 paradigm. *Allergy*. 2013;68(8):974-82.
- Bolognia J et al, "Basic Principles of Dermatology," Callen et al (eds.), *Dermatology (2017) 4th edition*, China: Elsevier.
- Bolognia J et al, "Atopic Dermatitis," Callen et al (eds.), *Dermatology (2017) 4th edition*, China: Elsevier.
- Thyssen JP, Kezic S. Causes of epidermal filaggrin reduction and their role in the pathogenesis of atopic dermatitis. *J Allergy Clin Immunol*. 2014;134(4):792-9.
- Cabanillas B, Novak N. Atopic dermatitis and filaggrin. *Curr Opin Immunol*. 2016;42:1-8.
- Kong HH et al. Temporal shifts in the skin microbiome associated with disease flares and treatment in children with atopic dermatitis. *Genome Res*. 2012;22(5):850-9.
- Williams MR et al. Quorum sensing between bacterial species on the skin protects against epidermal injury in atopic dermatitis. *Sci Transl Med*. 2019;11:490.
- Suárez-Fariñas M et al. Nonlesional atopic dermatitis skin is characterized by broad terminal differentiation defects and variable immune abnormalities. *J Allergy Clin Immunol*. 2011;127(4):954-64.
- Hijnen D et al. CD8(+) T cells in the lesional skin of atopic dermatitis and psoriasis patients are an important source of IFN- γ , IL-13, IL-17, and IL-22. *J Invest Dermatol*. 2013;133(4):973-9.
- Gittler JK et al. Progressive activation of T(H)2/T(H)22 cytokines and selective epidermal proteins characterizes acute and chronic atopic dermatitis. *J Allergy Clin Immunol*. 2012;130(6):1344-54.
- Pellerin L et al. Defects of filaggrin-like proteins in both lesional and nonlesional atopic skin. *J Allergy Clin Immunol*. 2013;131(4):1094-102.
- Danso MO et al. TNF- α and Th2 cytokines induce atopic dermatitis-like features on epidermal differentiation proteins and stratum

- corneum lipids in human skin equivalents. *J Invest Dermatol.* 2014;134(7):1941-50.
49. Hönzke S et al. Influence of Th2 cytokines on the cornified envelope, tight junction proteins, and β -defensins in filaggrin-deficient skin equivalents. *J Invest Dermatol.* 2016;136(3):631-9.
50. Papi A et al. Asthma. *Lancet.* 2018;391(10122):783-800.
51. Samitas K et al. Anti-IgE treatment, airway inflammation and remodelling in severe allergic asthma: Current knowledge and future perspectives. *Eur Respir Rev.* 2015;24(138):594-601.
52. Dullaers M et al. The who, where, and when of IgE in allergic airway disease. *J Allergy Clin Immunol.* 2012;129(3):635-45.
53. Säfholm J et al. Prostaglandin E2 inhibits mast cell-dependent bronchoconstriction in human small airways through the E prostanoid subtype 2 receptor. *J Allergy Clin Immunol.* 2015;136(5):1232-9.
54. Brown HM. Treatment of chronic asthma with prednisolone; significance of eosinophils in the sputum. *Lancet.* 1958;2(7059):1245-7.
55. Green RH et al. Asthma exacerbations and sputum eosinophil counts: A randomised controlled trial. *Lancet.* 2002;360(9347):1715-21.
56. Malinovschi A et al. Simultaneously increased fraction of exhaled nitric oxide levels and blood eosinophil counts relate to increased asthma morbidity. *J Allergy Clin Immunol.* 2016;138(5):1301-8.
57. Mogensen I et al. Simultaneously elevated FeNO and blood eosinophils relate to asthma morbidity in asthmatics from NHANES 2007-12. *Clin Exp Allergy.* 2018;48(8):935-43.
58. Dahlén B et al. Effect of formoterol with or without budesonide in repeated low-dose allergen challenge. *Eur Respir J.* 2009;33(4):747-53.
59. Rabe KF et al. Efficacy and safety of dupilumab in glucocorticoid-dependent severe asthma. *N Engl J Med.* 2018;378(26):2475-85.
60. Shaw DE et al. Clinical and inflammatory characteristics of the European U-BIOPRED adult severe asthma cohort. *Eur Respir J.* 2015;46(5):1308-21.
61. Balgoma D et al. Quantification of lipid mediator metabolites in human urine from asthma patients by electrospray ionization mass spectrometry: Controlling matrix effects. *Anal Chem.* 2013;85(16):7866-74.
62. Gómez C et al. Quantitative metabolic profiling of urinary eicosanoids for clinical phenotyping. *J Lipid Res.* 2019;60(6):1164-73.
63. Maric J et al. Cytokine-induced endogenous production of prostaglandin D2 is essential for human group 2 innate lymphoid cell activation. *J Allergy Clin Immunol.* 2019;143(6):2202-14.
64. Sanofi Genzyme and Regeneron. Type 2 innovation grant 2019. 2019. Available at: https://www.type2innovationgrant.com/prog/type_2_innovation_grant_2019/. Last accessed: 22 October 2019.

FOR REPRINT QUERIES PLEASE CONTACT: +44 (0) 1245 334450